Quantification of Smoking-Induced Occupancy of β2-Nicotinic Acetylcholine Receptors: Estimation of Nondisplaceable Binding

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5-123I-iodo-85380 (123I-5-IA) is used to quantitate high-affinity nicotinic acetylcholine receptors (β2-nAChRs) on human SPECT scans. The primary outcome measure is \( V_i/f_p \), the ratio of equilibrium between total tissue concentration (free, nonspecifically bound, and specifically bound) and the free plasma concentration. Nondisplaceable uptake (free plus nonspecific) of 123I-5-IA has not been measured in human subjects. Nicotine has high affinity for β2*-nAChRs (nAChRs containing the β2* subunit, for which * represents other subunits that may also be part of the receptor) and displaces specifically bound 123I-5-IA. In this study, we measured nicotine occupancy and nondisplaceable binding in healthy smokers after they had smoked to satiety.

Methods: Eleven nicotine-dependent smokers (mean age ± SD, 35.6 ± 14.4 y) completed the study. One subject was excluded from subsequent analyses because of abnormal blood nicotine levels. Subjects abstained from tobacco smoke for 0.9 d and participated in a 15- to 17-h SPECT scanning day. 123I-5-IA was administered by bolus plus constant infusion, with a total injected dose of 361 ± 20 MBq. At approximately 6 h after the start of the infusion, three 30-min SPECT scans and a 15-min transmission-emission scan were acquired to obtain baseline β2*-nAChR availability. Subjects then smoked to satiety (2.4 ± 0.7 cigarettes), and arterial (first 40 min) and venous (until study completion) plasma nicotine and cotinine levels were collected. About 1 h after subjects had smoked to satiety, up to six 30-min SPECT scans were acquired. \( V_i/f_p \) data, computed from the tissue and plasma radioactivity measurements from the presmoking baseline and postsmoking scans, were analyzed using the Lassen plot method.

Results: Receptor occupancy after subjects had smoked to satiety was 67% ± 9% (range, 55%–80%). Nondisplaceable uptake was estimated as 19.4 ± 5.8 mL/cm³ (range, 15–28 mL/cm³). Thus, in the thalamus, where mean \( V_i/f_p \) is 93 mL/cm³, nondisplaceable binding represents approximately 20% of the total binding.

Conclusion: These results are in agreement with previous findings and suggest that when satiating doses of nicotine are administered to smokers, imaging of receptor availability can yield valuable data, such as quantifiable measures of nondisplaceable binding.

Key Words: SPECT; 5-123I-iodo-85380; smoking to satiety; human smokers

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Nicotinic acetylcholine receptors (nAChRs) are abundant throughout the peripheral and central nervous systems, are the primary targets for nicotine in the brain, and play an important role in cognition and behavior. The nAChRs are pentameric ligand–gated ion channels. Many of the subunits have been identified, but nicotine has the highest affinity for nAChRs containing the β2*-subunit (β2*-nAChRs, for which * represents other subunits that may also be part of the receptor), which are the most widely expressed in the brain. Changes in ligand binding to the high-affinity state of β2*-nAChRs have been demonstrated in tobacco smoking (1,2) and psychiatric and neurocognitive disorders including schizophrenia (3,4) and Alzheimer disease (5,6). In vivo imaging of β2*-nAChRs has significantly advanced since the development of 3-[2(S)-2-azetidinylmethoxy]pyridine (A-85380) (7), which is used for SPECT when labeled with 123I (8) or for PET when labeled with 18F (9,10). A-85380 has a high affinity and low dissociation from the receptor–ligand complex, high specific-to-nonspecific binding ratio, and high selectivity for the β2*-nAChR (8,11). Consistent with the known distribution of β2*-nAChRs, A-85380 displays a high binding in the thalamus, an intermediate binding in the cortex and striatum (12–14), and the lowest binding in the cerebellum (14,15). 5-123I-iodo-85380 (123I-5-IA) is a halogenated analog of A-85380 (16) and is safe and nontoxic (17,18). Further, there is good agreement between bolus and bolus plus infusion paradigms using 123I-5IA, indicating that quantitation of β2*-nAChR...
availability using this ligand is accurate (19). Use of A-85380 has been verified in animal (11,19,20) and human (17,21,22) studies.

Our group has quantified β_{2*-nAChR} availability, measured with 123I-5-IA SPECT, using V_{T}/f_{p}—which represents regional brain concentration (kBq/cm³) divided by free parent concentration in plasma (f_{p}; kBq/mL) (23,24) and equals the sum of specific (V_{S}/f_{p}) and nondisplaceable (V_{ND}/f_{p}) binding. Specifically, we demonstrated significant individual variability in protein binding (23); observed sex differences in the volume of distribution were due to differences in protein binding of the radiotracer. Thus, it is critical to correct for free fraction (f_{p}) in our outcome measure and report V_{T}/f_{p} as opposed to V_{T}. However, by reporting V_{T}/f_{p}, we do not correct for V_{ND}/f_{p}. To obtain a more precise measure of β_{2*-nAChR} availability in vivo, an accurate measurement of nonspecific binding has to be made. The purpose of the present study was to estimate V_{ND}/f_{p} of 123I-5-IA in human tobacco smokers by examining displacement through saturation of receptors by the amount of tobacco smoke required to reach satiety and applying a graphical analysis of the V_{T}/f_{p} change from baseline to estimate both occupancy and V_{ND}/f_{p}. In individuals who smoke, this can be done safely through self-administration of tobacco cigarettes to satiating levels (25). This study was conducted in smokers rather than nonsmokers so that participants could safely self-administer satiating levels of nicotine without adverse side effects such as the significant increase in blood pressure and disorientation (26), increased tension, confusion, and decreased vigor (27,28) that can occur in nicotine-naïve individuals after administration of a small dose of nicotine (such as from a nicotine patch). We calculated V_{ND}/f_{p} after smokers smoked to satiety, which has not been previously reported. We also examined associations between receptor occupancy and smoking parameters and hypothesized that greater receptor occupancy would be reached through smoking a greater number of cigarettes.

**MATERIALS AND METHODS**

**Participants**

Eleven volunteers (5 men and 6 women; mean age ± SD, 35.6 ± 14.4 y), who smoked 17.5 ± 6.8 cigarettes/d for 19.6 ± 14.2 y (Table 1), participated in the study. Nine subjects were Caucasian, 1 African American, and 1 Asian American. They provided written informed consent to participate in this study, conducted at Yale University and the West Haven Veteran’s Affairs Connecticut Healthcare System; the study was approved by the Human Investigational Research Committees at both institutions. Eligibility was evaluated via structured interview, behavioral assessments, physical examination, laboratory blood tests, urine drug screen, and an electrocardiogram. None of the subjects had a recent history or evidence of a serious medical or neurologic illness, head trauma with significant loss of consciousness, psychiatric disorder, or substance abuse (except for nicotine dependence) as determined by the Structured Clinical Interview for the Diagnostic and Statistical Manual of Mental Disorders, 4th edition (29). Subjects had not used psychotropic substances other than alcohol for at least 1 y and had not used marijuana for at least 1 mo preceding the study. Women were not pregnant or breast-feeding. Nicotine dependence and current depressive symptoms were evaluated with the Fagerstrom Test of Nicotine Dependence, Center for Epidemiological Studies Depression Scale, and Beck Depression Inventory at intake and on the day of scanning.

**Smoking Cessation**

Two weeks before the SPECT scanning day, subjects met with research staff to learn tobacco smoking—abstinence techniques and set the date for smoking cessation. All subjects were asked to abstain from smoking for 4–6 d (mean ± SD, 5.3 ± 0.9 d) before the SPECT scanning day to allow time for residual nicotine to clear from the brain (30). Nonsmoking status was confirmed by breath carbon monoxide levels less than 11 pulses per minute and negligible plasma (≤100 ng/mL) and urinary (<200 ng/mL) cotinine levels on the scanning day.

**MRI and 123I-5-IA SPECT**

**MRI.** Each subject participated in 1 MRI scan before SPECT. MRI was performed on a Sigma 1.5-T system (GE Healthcare). Axial images parallel to the anteroposterior commissural line were acquired with the following parameters: echo time, 5 ms; repetition time, 24 ms; matrix, 256 × 192; number of excitations, 1; field of view, 24 cm; and 128 contiguous slices with 1.3-mm thickness.

**SPECT.** All emission scans were obtained on a Prism 3000 XP (Picker), a 3-head SPECT camera equipped with a low-energy, ultra-high-resolution fanbeam collimator (photopeak window, 159 keV ± 10%); matrix, 128 × 128) with a uniform sensitivity across the field of view. A 57Co-distributed source was measured with each experiment to control for day-to-day variation in camera sensitivity. The axial resolution (full width at half maximum) was 12.2 mm, measured with a 123I line source in water in a cylindrical phantom. 123I-5-IA was synthesized as previously described (31) and administered through a venous catheter placed in the arm or hand using a bolus plus constant infusion with a ratio of 7.0 ± 0.02 h and a total injected dose (accounting for decay) of 361 ± 20 MBq. Another venous catheter was placed into the opposite arm or hand to collect blood for protein binding and metabolism. Previous data show that after 2 h of 5-IA infusion, venous and arterial samplings of the radiotracer are comparable (32). Therefore, we use venous sampling to obtain radiotracer protein binding and metabolism during the scanning period. Approximately 6 h after injection of 123I-5-IA, a simultaneous transmission—emission scan and 3 equilibrium emission scans were obtained. Subjects were then removed from the camera, and a line was placed in the radial artery by an anesthesiologist after the Allen test was performed to assess collateral ulnar flow. An arterial catheter was used to collect blood for analysis of nicotine and its metabolite cotinine. The subjects were instructed to smoke up to 5 cigarettes of their preferred brand within 20 min, through a smoking topography device (Plowshare Technologies, Borgwaldt KC, Inc.) to control for any difference in puff volume. Because initial nicotine levels in arterial blood are higher than in venous samples (33), we collected arterial blood samples for the first 40 min of cigarette smoking, starting at the time of inhalation of the first cigarette, to determine nicotine and cotinine levels, after which venous samples were collected hourly until study completion. After the smoking challenge, one 30-min emission scan was acquired every hour for up to 6 h (all but 1 subject participated in all 6
scans) to evaluate displacement of $^{123}$I-5-IA from $\beta_2^\ast$-nAChRs throughout the brain. Blood samples were collected at the midpoint of each postsmoking scan to quantify total parent and $f_p$ to correct for individual differences in metabolism and protein binding of $^{123}$I-5-IA.

**Image Analysis**

SPECT emission images were analyzed as described previously (21). SPECT emissions were filtered using a 3-dimensional Butterworth filter (power factor, 10; cutoff, 0.24 cycle/pixel) and reconstructed using a filtered backprojection algorithm with a ramp filter on a 128 × 128 matrix to obtain 50 slices with a pixel size of 2.06 × 2.06 × 3.56 mm in the $x$, $y$, and $z$-axes. An attenuation map was reconstructed from the transmission and flood data, and nonuniform attenuation correction was performed (34). MR images were coregistered to each SPECT image to provide an anatomic guide for placement of standardized 2-dimensional regions of interest using MEDx software (Medical Numerics, Inc.). A 3-dimensional volume of interest was generated for each region and transferred to the coregistered SPECT image to determine regional radioactivity concentrations. The chosen regions of interest were those known to contain $\beta_2^\ast$-nAChRs: the frontal, parietal, anterior cingulate, temporal, and occipital cortices; thalamus; striatum (an average of caudate and putamen); and cerebellum. Regional $^{123}$I-5-IA uptake was determined by $\frac{V_T}{f_p}$, where $V_T$ is brain regional activity divided by metabolite-corrected plasma activity and $f_p$ is plasma parent. This measure is highly reproducible (21) and was previously designated as $V_T$, but the nomenclature was changed on the basis of a study by Innis et al. (24). Interrater variability, which was less than 10% across all regions, was computed as percentage difference between 2 raters using the equation ($\frac{V_T1}{V_T2} - 1) \times 100$.

**TABLE 1. Demographic Variables and SPECT Scan Smoking Outcomes**

<table>
<thead>
<tr>
<th>Cigarette</th>
<th>Subject no.</th>
<th>Age (y)</th>
<th>Race</th>
<th>Sex</th>
<th>Preferred brand</th>
<th>Amount of nicotine (mg)</th>
<th>FTND</th>
<th>Days abstinent</th>
<th>Cmax nicotine (ng/mL)</th>
<th>Tmax nicotine (min)</th>
<th>Cmax nicotine (mg/mL)</th>
<th>f_p</th>
<th>Cmax nicotine (mg/mL)</th>
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<td>1</td>
<td>22</td>
<td>36 ± 15</td>
<td>Asian</td>
<td>Female</td>
<td>Parliament</td>
<td>0.7</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>20.0</td>
<td>27.3</td>
<td>8,544.3 ± 7,068.8</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>38</td>
<td>5.3 ± 14.0</td>
<td>Caucasian</td>
<td>Male</td>
<td>Marlboro</td>
<td>1.0</td>
<td>4.0</td>
<td>2.0</td>
<td>4.0</td>
<td>4.0</td>
<td>63.0</td>
<td>7,153.0</td>
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<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>2.0</td>
<td>45.0</td>
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</tr>
<tr>
<td>4</td>
<td>50</td>
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<td>Female</td>
<td>Virginia Slims</td>
<td>0.7</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>2.0</td>
<td>45.0</td>
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<td>Male</td>
<td>Marlboro</td>
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<tr>
<td>6</td>
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<td>Female</td>
<td>Virginia Slims</td>
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<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>2.0</td>
<td>50.0</td>
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<td></td>
</tr>
<tr>
<td>7</td>
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<td>Male</td>
<td>Newport</td>
<td>1.2</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>2.0</td>
<td>32.4</td>
<td>8,544.3 ± 7,068.8</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>48</td>
<td>4.0</td>
<td>Caucasian</td>
<td>Male</td>
<td>Newport</td>
<td>1.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>2.0</td>
<td>20.0</td>
<td>27.3</td>
<td>8,544.3 ± 7,068.8</td>
</tr>
<tr>
<td>9</td>
<td>39</td>
<td>0.7</td>
<td>Caucasian</td>
<td>Female</td>
<td>Parliament</td>
<td>1.0</td>
<td>4.0</td>
<td>4.0</td>
<td>4.0</td>
<td>2.0</td>
<td>20.0</td>
<td>27.3</td>
<td>8,544.3 ± 7,068.8</td>
</tr>
<tr>
<td>10</td>
<td>20</td>
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<td>Hispanic</td>
<td>Male</td>
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<td>4.0</td>
<td>4.0</td>
<td>2.0</td>
<td>20.0</td>
<td>27.3</td>
<td>8,544.3 ± 7,068.8</td>
</tr>
</tbody>
</table>

**FIGURE 1.** $\beta_2^\ast$-nAChR occupancy plot based on Lassen method for representative subject’s scan. Receptor occupancy by nicotine is determined from slope of linear regression ($\theta$), 74.4% in this case. $V_{ND/f_p}$ is calculated as $y$-intercept divided by slope. For this scan, $V_{ND/f_p} = 16.5$ mL·cm$^{-3}$. 

where $\frac{V_T}{f_p}$ is the ratio of brain regional activity to metabolite-corrected plasma activity. $V_{ND/f_p}$ is calculated as the $y$-intercept divided by the slope of the linear regression of $\frac{V_T}{f_p}$ versus $\frac{V_T}{f_p}$. This measure is highly reproducible (21) and was previously designated as $V_T$, but the nomenclature was changed on the basis of a study by Innis et al. (24). Interrater variability, which was less than 10% across all regions, was computed as percentage difference between 2 raters using the equation ($\frac{V_T1}{V_T2} - 1) \times 100$. 

**Determination of Receptor Occupancy and $V_{ND/f_p}$**

$V_{ND/f_p}$ data, computed directly from the tissue and plasma radioactivity measurements from the presmoking baseline and postsmoking scans, were analyzed using Lassen plots. These plots have been validated by a recent study (35), with the equation for the line ($y = rx + b$), where $y = \frac{V_T}{f_p}$ [baseline] - $V_{ND/f_p}$ [after nicotine], $x = V_T$ (baseline), and linear regression of the fit to different brain regions provides receptor occupancy ($\theta$) and $V_{ND/f_p}$. 

\[
y = 0.7443x - 12.304
\]
Receptor occupancy and derivation of $V_r$ between receptor occupancy and $f_p$ were thus derived for each subject across all brain regions for each postsmoking scan (compared with the presmoking baseline), and the final result represents the average across scans for each subject. We also derived $V_{ND/f_p}$ without correcting for $f_p$ to examine the role $f_p$ plays in the derivation of $V_{ND/f_p}$.

**Statistical Analysis**

All statistical analyses were performed using SPSS (version 15.0; SPSS Inc.). Nonparametric correlational analyses (Spearman $r$-correlation coefficient) were used to examine the relationship between receptor occupancy and $V_{ND/f_p}$ variables and smoking status. Statistical significance was set at $P$ less than or equal to 0.05.

**RESULTS**

Mean total scores ± SD for the Fagerstrom Test of Nicotine Dependence (5.3 ± 2.8), Beck Depression Inventory (1.7 ± 1.7), and Center for Epidemiological Studies Depression Scale (4.8 ± 4.3) indicated that subjects were moderately dependent on tobacco and had negligible symptoms of depression during the study. At the time of screening, subjects had exhaled carbon monoxide levels (18.6 ± 6.1 pulses per minute) and plasma cotinine levels (228.2 ± 155.1 ng/mL) consistent with tobacco dependence. On the SPECT scanning day, subjects had low exhaled carbon monoxide (3.3 ± 3.8 pulses per minute) and plasma cotinine (55.0 ± 41.7 ng/mL) levels consistent with abstinence, as required for participation. After the set of baseline equilibrium scans, subjects smoked to satiety (mean cigarettes smoked ± SD, 2.4 ± 0.7). One subject showed abnormal nicotine metabolism after the smoking challenge. The typical pattern for nicotine availability in blood is to peak at approximately 10–20 min after smoke inhalation and subsequently to decrease over the next 1–2 h, depending on the amount of smoke inhalation and individual metabolism. On the basis of typical nicotine kinetics, we expected nicotine plasma levels to drop below detection levels (4 ng/mL) within 2–3 h after the smoking challenge. However, the excluded subject’s nicotine levels in blood continued to rise through study completion, up to 6 h after cigarette smoking. This subject was excluded from subsequent analyses. For the rest of the subjects (1–10), after the smoking challenge the peak in nicotine levels was reached at 8.1 ± 6.9 min (Table 1) and maximum nicotine levels were 51.4 ± 24.9 ng/mL (Fig. 2B, representative subject).

$f_p$

The group mean and SD for $f_p$ was 0.10 ± 0.04 (range, 0.6–1.6). Individual $f_p$ values were used in subsequent calculations unless otherwise noted.

**Receptor Occupancy**

Equilibrium, operationally defined as less than a 5% change in receptor availability per hour (mL·cm$^{-3}$·h$^{-1}$), was reached between 6 and 8 h after injection (Fig. 2; representative subject, average change was 3.1%/h in the thalamus, 3.9%/h in the striatum, 0.1%/h in the mean cortex, and 2.7%/h in the cerebellum). After the smoking challenge (i.e., 2–5 h after smoking), there was minimal change in radiotracer binding (Table 2; Fig. 3), suggesting a pseudoequilibrium was reached, similar to a minimal change in binding a reported previously (3.1 h after smoking challenge) (25).

Peak occupancy of 69.1% ± 9.1% was reached at 3 h after the challenge. At 1 h after the challenge (i.e., 1 h from the start of smoking), maximum occupancy had not been reached, and at 6 h after the challenge, the receptor occupancy by nicotine started to decline (Table 2). Therefore, the data from scans 1 and 6 are not used in the derivation of final occupancy parameters. The decrease in $\beta_2$*-nAChR availability (or receptor occupancy by nicotine) after nicotine challenge was dramatic (Fig. 4; representative subject illustrated), with a maximal occupancy of 69.1% at 3 h after the smoking challenge and a range in receptor occupancy of 57.6%–69.1% over the 5–h scanning period for this subject. In the overall sample, the displacement of radiotracer by nicotine was 55%–80% across subjects and scans, with an average occupancy.

**FIGURE 2.** Time–activity curve (A) and associated blood plasma nicotine concentration (B) from representative subject. (A) Equilibrium was reached before subject had smoked to satiety: for this subject, average percentage change was 3.1%/h in thalamus, 3.9%/h in striatum, 0.1%/h in mean cortex, and 2.7%/h in cerebellum. Two to 5 h after smoker had smoked to satiety, there appeared to be little change in kBq/cm$^3$, suggesting that pseudoequilibrium state was achieved. (B) Highest nicotine concentration was observed at 20 min after initiation of tobacco smoking.
across scans 2–5 (2–5 h after the challenge) of 67.2% ± 9.1%.

\[ V_{ND/FP} \]

The estimates of \( V_{ND/FP} \) remained stable over the course of the postsmoking challenge (Fig. 5), ranging from 15% to 28% across subjects, with an average of 19.4 ± 5.8 mL/cm\(^3\) (SD of 27.6%) for \( V_{ND/FP} \). Thus, because baseline \( V_{T/FP} \) in the thalamus was 93.5 mL/cm\(^3\), \( V_{ND/FP} \) represents 20.7% of the activity in this region.

We also estimated nondisplaceable binding without correcting for \( f_p \). Average \( V_{ND} \) was 7.03 ± 1.92 kBq/cm\(^3\), and SD was 26.3%. Therefore, it does not appear that correction for plasma free fraction reduced the variability of nondisplaceable binding in this sample.

**Data Correlations**

We examined if smoking variables play a role in receptor occupancy by nicotine or \( V_{ND/FP} \) by conducting a nonparametric correlation analysis (Spearman \( r \)). We observed a positive correlation between number of cigarettes smoked on the scanning day (cigarette challenge to satiety) and receptor occupancy at hours 5 (\( r = 0.67, P = 0.05 \)) and 6 (\( r = 0.73, P = 0.03 \)) after challenge only (Fig. 6). There was no significant correlation between the number of cigarettes smoked on the scanning day and the average occupancy for hours 1–6 of scanning.

**DISCUSSION**

In the current study, we calculated \( \beta_2^-\text{nAChR} \) occupancy by nicotine by having human tobacco smokers smoke to satiety. Instead of saturating receptors with nicotine through alternate methods such as intravenous nicotine, having subjects smoke to satiety provides a more accurate quantification of receptor occupancy at the times that individuals are satisfied with their use of tobacco smoke. Consistent with a previous study (25), we demonstrated that in a typical tobacco smoker, most of the receptors remain occupied for a prolonged period. Specifically, in the current study, smoking an average of 2.4 cigarettes occupied up to 80% of receptors for up to 5 h; these results are comparable but somewhat lower than the previous report by Brody et al. (I), in which the receptor occupancy was 95% after subjects had smoked to satiety (2.8 cigarettes). One potential reason for this difference may be the method by which receptor occupancy was calculated. In the study by Brody et al., percentage occupancy was calculated as a ratio of receptors occupied after the challenge to receptors occupied before the challenge in each brain region and corrected for the amount of nicotine present in the subject’s blood at the time of greatest occupancy (3.1 h)—typically less than 4 ng/mL. However, because there is no physiologic reason to assume differences in receptor occupancy across regions, we used the well-known Lassen method (35), previously used to calculate receptor occupancy. Further, because the lower limit of detection of nicotine in blood in our laboratory is 4 ng/mL and the half-life of nicotine is about 2.5 h, we were not able to correct for the amount of nicotine present in blood at the time of maximal occupancy.

As expected and consistent with a previous study (25), \( \beta_2^-\text{nAChR} \) occupancy was related to the number of cigarettes smoked on the scanning day (smoking-to-satiety challenge). We detected a significant positive correlation between receptor occupancy and the number of cigarettes smoked at challenge hours 5 and 6 of scanning only, suggesting that greater receptor occupancy was achieved by smoking a greater number of cigarettes.

We also determined the magnitude of \( V_{ND/FP} \) (sum of nonspecific and free) for \( ^{123}\text{I-5-IA} \) for the first time, to our knowledge, in human subjects. This determination was

**TABLE 2. Percentage Occupancy for Each Subject at 1–6 Hours After Smoking**

<table>
<thead>
<tr>
<th>Subject no.</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>Mean SD</th>
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<td>63.7</td>
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<td>N/A</td>
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<td>79.5</td>
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<td>79.9</td>
<td>79.1</td>
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<td>60.9</td>
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**N/A = not applicable. Italized numbers are used to calculate final average receptor occupancy. Mean of receptor occupancy for scans between hours 2 and 5 after smoking challenge was used.**
made by having tobacco smokers smoke to satiety (36). In addition, the current study was conducted exclusively in subjects who were habitual smokers because administration of saturating doses of nicotine to nicotine-naïve subjects can have serious side effects and also raises the ethical issue of giving drugs to drug-naïve individuals (37). We determined that, at baseline, $V_{ND/fp}$ represents about 20% of total $^{123}$I-5-IA binding in the thalamus, the mostly densely populated $\beta_2$*-nAChR region. We observed between-subject variability in $V_{ND/fp}$, which is likely due to individual differences in nonspecific binding and free radiotracer concentration in the brain as measured with $^{123}$I-5-IA SPECT. We did not observe significant variability in $V_{ND/fp}$ across the 6-h scanning period, suggesting that the full 6-h block is not necessary to measure $V_{ND/fp}$. However, here we used the average across all 6 scans to report mean group $V_{ND/fp}$ that may be used to obtain a more precise measure of receptor availability in future studies.

Further, although determining the magnitude of nondisplaceable binding for $^{123}$I-5-IA correction for $f_p$ may not be necessary, because of population differences in $f_p$, this correction is important. Previous reports in nonhuman primates estimated specific binding to be at about 70% in the thalamus; thus, nondisplaceable binding (specific and free) would be about 30%, which is slightly higher than the percentage in the current sample (19). The current results will allow future studies to determine $\beta_2$*-nAChR availability more accurately.

There are several limitations to the current study. First, the finite dissociation and brain clearance of $^{123}$I-5-IA from the receptor–ligand complex limits the use of $^{123}$I-5-IA, or any radioligand, and neuroreceptor imaging techniques (8,11,38). Finite dissociation and brain clearance mean that the radioligand binding to the receptor does not instantaneously match the available receptor. Further, even if the ligand dissociates from the receptor because of competition (e.g., from drug), this tracer must leave the brain to detect a change in the imaging signal. In the case of nicotine, tobacco smoke enters the body through the mouth, is inhaled into the lungs, and is then absorbed into the circulatory system that delivers it to the brain. When delivered this way to dependent smokers, nicotine from tobacco smoke reaches the brain within approximately 30 s of inhalation (39). A typical tobacco smoker requires approximately 5 min to smoke a cigarette. With nicotine’s half-life of approximately 90 s in the lungs, and with the brain uptake being highly correlated with rate of washout from the lung (39), it is probable that the maximal brain concentration and receptor occupancy are reached much earlier than measured by $^{123}$I-5-IA, because it takes $^{123}$I-5-IA quite some time to dissociate from the receptor and exit the brain. In fact, by 3 h after smoking, the occupancy by nicotine may have dropped enough so that $^{123}$I-5-IA uptake levels begin to rebound.

A second potential limitation to studying the nAChRs using A-85380 is that endogenous acetylcholine competes...
with $^{123}$I-5-IA for binding to nAChRs (39). Fujita et al. (40) showed that $^{123}$I-5-IA was displaced by increased levels of acetylcholine, and the change in 5-IA levels was caused by a change in specific, not nonspecific, binding. This means that an increase in baseline levels of acetylcholine occupancy will decrease the binding of $^{123}$I-5-IA, a potentially relevant finding in the current study because some of the receptors may be occupied by acetylcholine and not available for nicotine or the radioligand. In this case, the baseline $V_{T/FP}$ values, and the magnitude of occupancy by nicotine, will be underestimated. The low variance between occupancy measures across subjects suggests that despite different numbers of cigarettes smoked, $\beta_2^*-nAChR$ occupancy is a reasonably good imaging biomarker, relevant to behavioral states in the subject. Third, these findings may not generalize to psychiatric populations. Though it is likely that there are no significant differences in $V_{NDFP}$ between populations, we cannot state with confidence that the $V_{NDFP}$ as reported here is appropriate to use in psychiatric populations without further study. We are currently evaluating $\beta_2^*-nAChR$ availability in psychiatric populations, including smokers with schizophrenia and alcohol-use disorders. Future work will need to examine whether $V_{NDFP}$ is uniform among psychiatric populations, as compared with healthy controls, to ensure that the differences in receptor binding are not due to differences in $V_{NDFP}$. Presently, the $V_{NDFP}$ value as calculated here may be used with caution when reporting $V_{T/FP}$ in psychiatric populations. Because $V_{NDFP}$ accounts for approximately 20% only of thalamic binding, a large between-group change in $V_{NDFP}$ would be needed to affect the interpretation of between-group differences in $V_{T/FP}$.

**CONCLUSION**

To date, no neuroimaging studies using $^{123}$I-5-IA report calculations of $V_{NDFP}$. Presently, we demonstrate that receptor occupancy by nicotine from satiating tobacco smoke levels reaches a maximum at 2–5 h after nicotine challenge, which is somewhat contradictory to smokers’ self-report of craving tobacco smoke every 1–2 h. We propose that one of the reasons for this discrepancy is the slow kinetics of $^{123}$I-5-IA and that the maximal receptor occupancy by nicotine after use of a cigarette might be reached more quickly. In future studies, we plan to examine the relationship between nicotine occupancy of $\beta_2^*-nAChR$ and tobacco smoke craving, urges, and withdrawal to better understand other aspects of cigarette smoking that may contribute to frequent daily use of cigarettes.

Further, we calculated nondisplaceable uptake of $^{123}$I-5-IA ($V_{NDFP}$) and demonstrated that it was approximately 20% of total binding for this radiotracer in the thalamus of healthy human smokers. This finding suggests that $^{123}$I-5-IA is an excellent tracer to continue the investigation of $\beta_2^*-nAChR$ in vivo. Further, this study provides a more precise measurement of $\beta_2^*-nAChR$ availability in vivo, and $\beta_2^*-nAChR$ should be considered in future studies using $^{123}$I-5-IA for a more accurate measure of $V_{T/FP}$.

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**REFERENCES**